



# L-Form Bacteria and their role in Antimicrobial Resistance

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# Outline

## Introduction

- L-form bacteria
- Urinary tract infections

## Potential role of L-form switching in AMR

- Isolation of L-forms in rUTI patients
- Visualization of L-form bacteria
- L-form switching is reliant on antibiotic presence

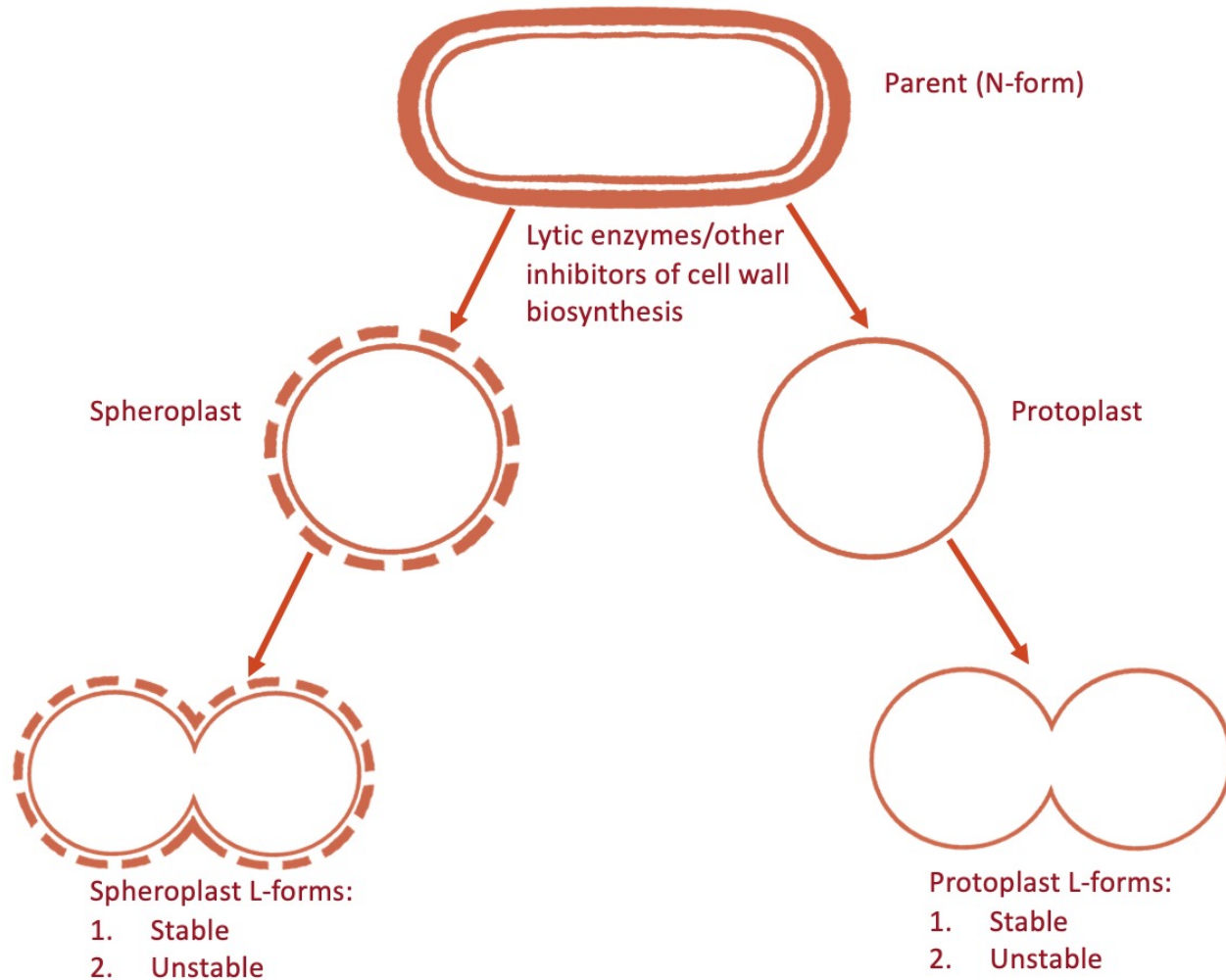
## Discussion

- Other bacterial pathogens with L-form mechanisms
- Reassessing routine antibiotics used for UTIs

## Conclusions and future directions

# L-Form Bacteria

- L-form/L-phase/**cell wall-deficient**
- Discovered in 1935 by Emmy Klieneberger-Nobel at the Lister Institute



# L-Forms as a novel Antimicrobial Resistance mechanism

- Cell-wall components recognized by immune system
- Target for immune effectors
- Target for antibiotics

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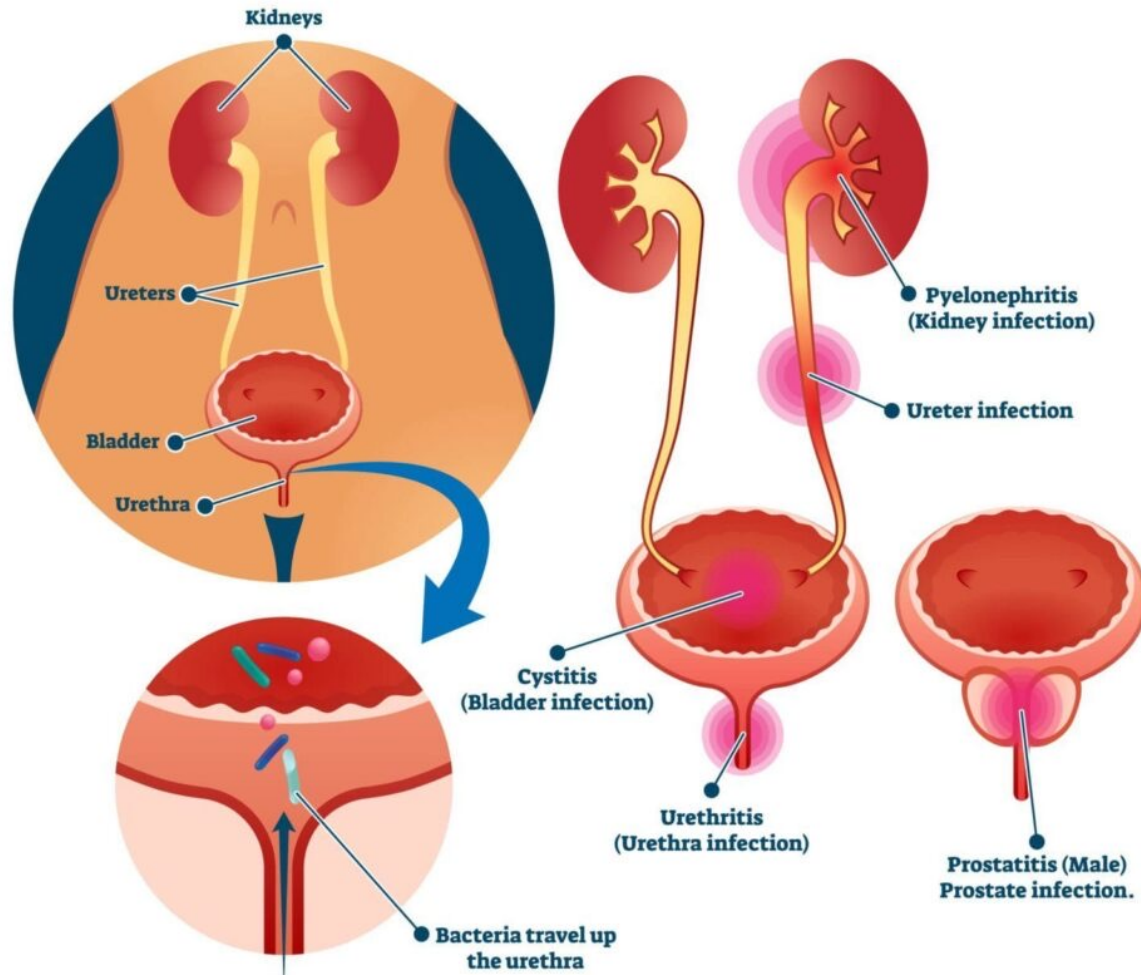
## Possible role of L-form switching in recurrent urinary tract infection

[Katarzyna M. Mickiewicz](#) ✉, [Yoshikazu Kawai](#), [Lauren Drage](#), [Margarida C. Gomes](#), [Frances Davison](#), [Robert Pickard](#), [Judith Hall](#), [Serge Mostowy](#), [Phillip D. Aldridge](#) & [Jeff Errington](#) ✉

[Nature Communications](#) **10**, Article number: 4379 (2019) | [Cite this article](#)

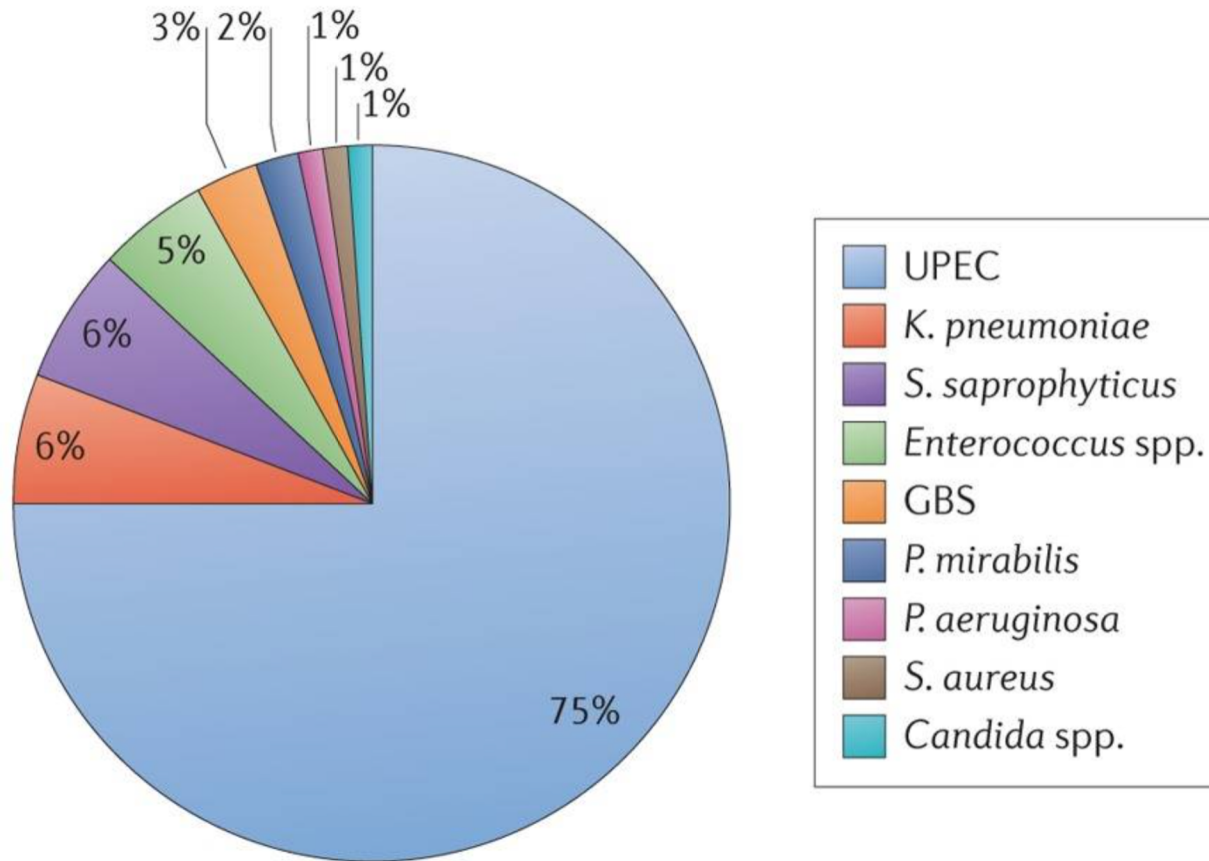
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# Urinary tract infection (UTI)

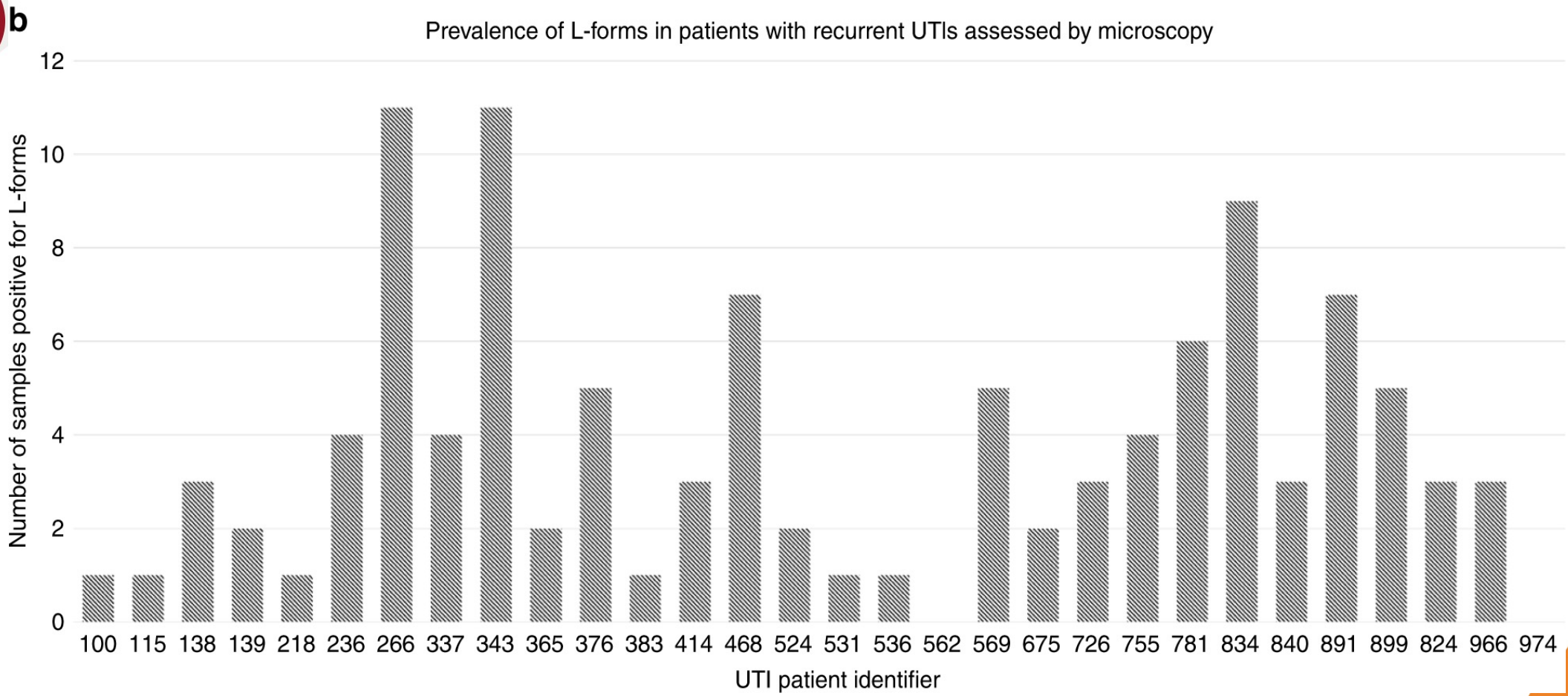


- Infection in any part of urinary system
- Most common outpatient infection
- 11% prevalence
  - 20% in >65 women
- 40-60% women will have a UTI in their lifetimes
  - 25% have rUTI
- Annual cost of **\$1.6 billion** in US

# Uropathogenic *Escherichia coli* (UPEC)

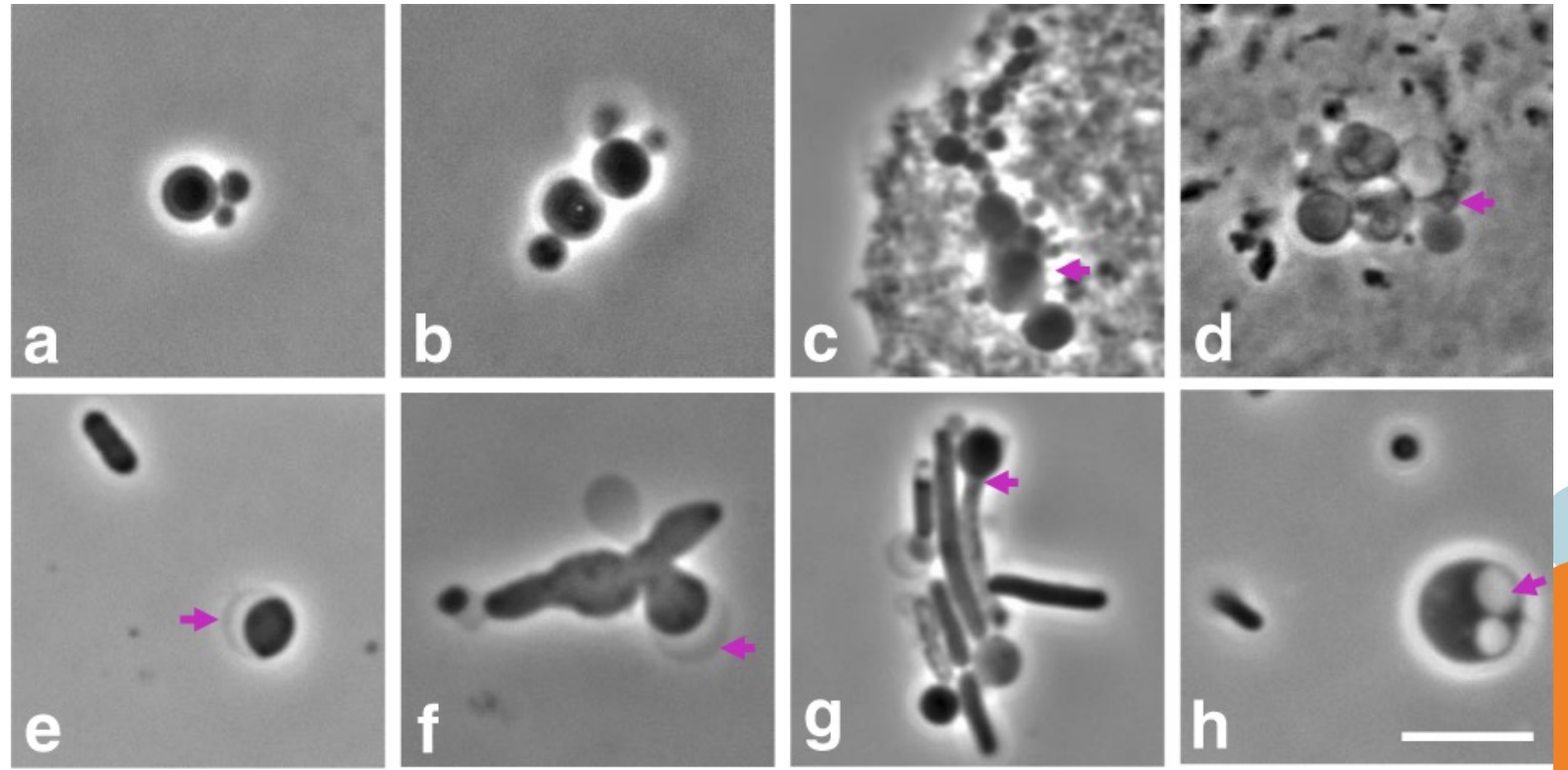


- Agents causing UTIs are commensal organisms
- 50-80% UTIs cause by *E. coli*



# L-Forms found in rUTI urine samples via phase contrast microscopy

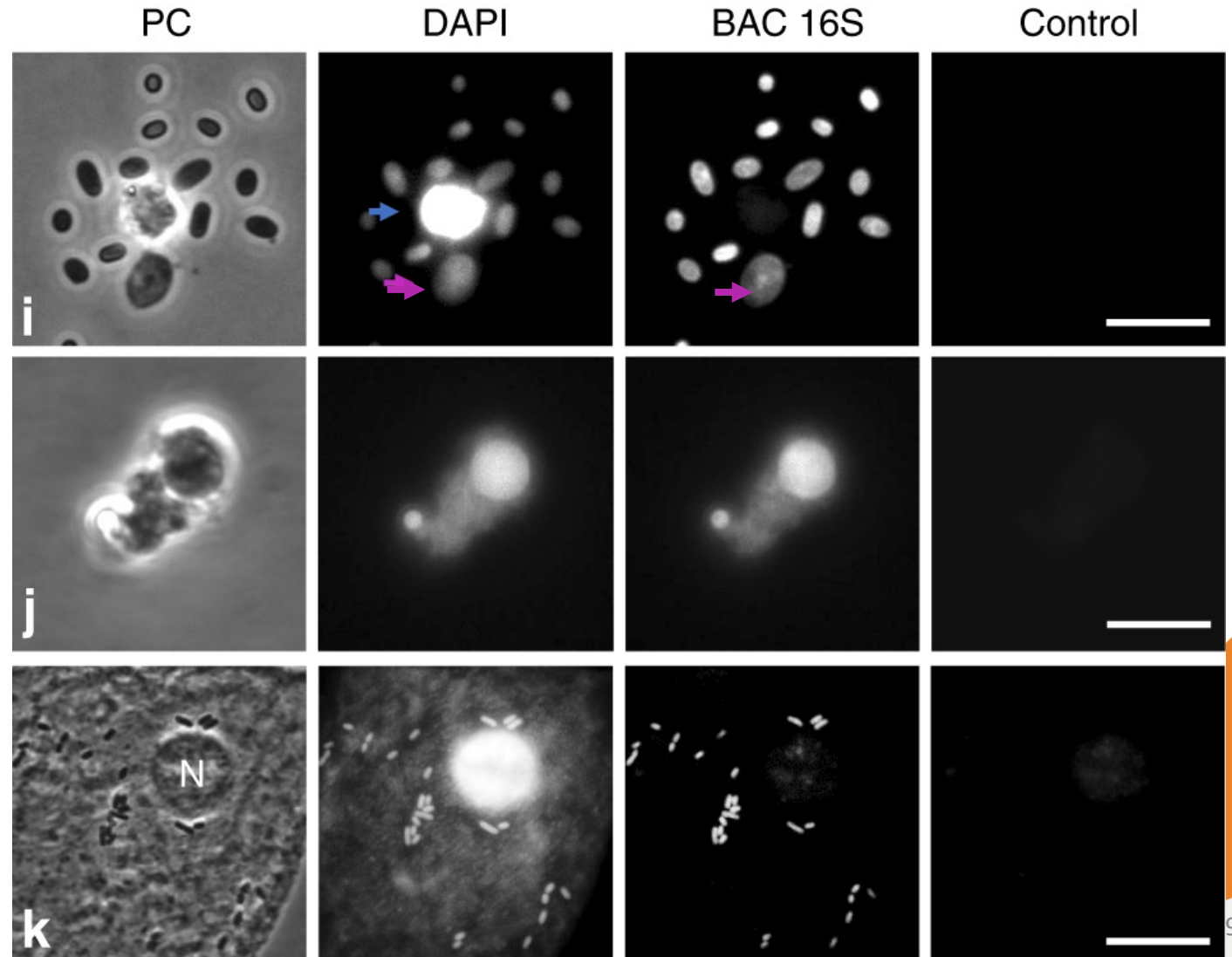
- Fresh urine samples phase contrast microscopy





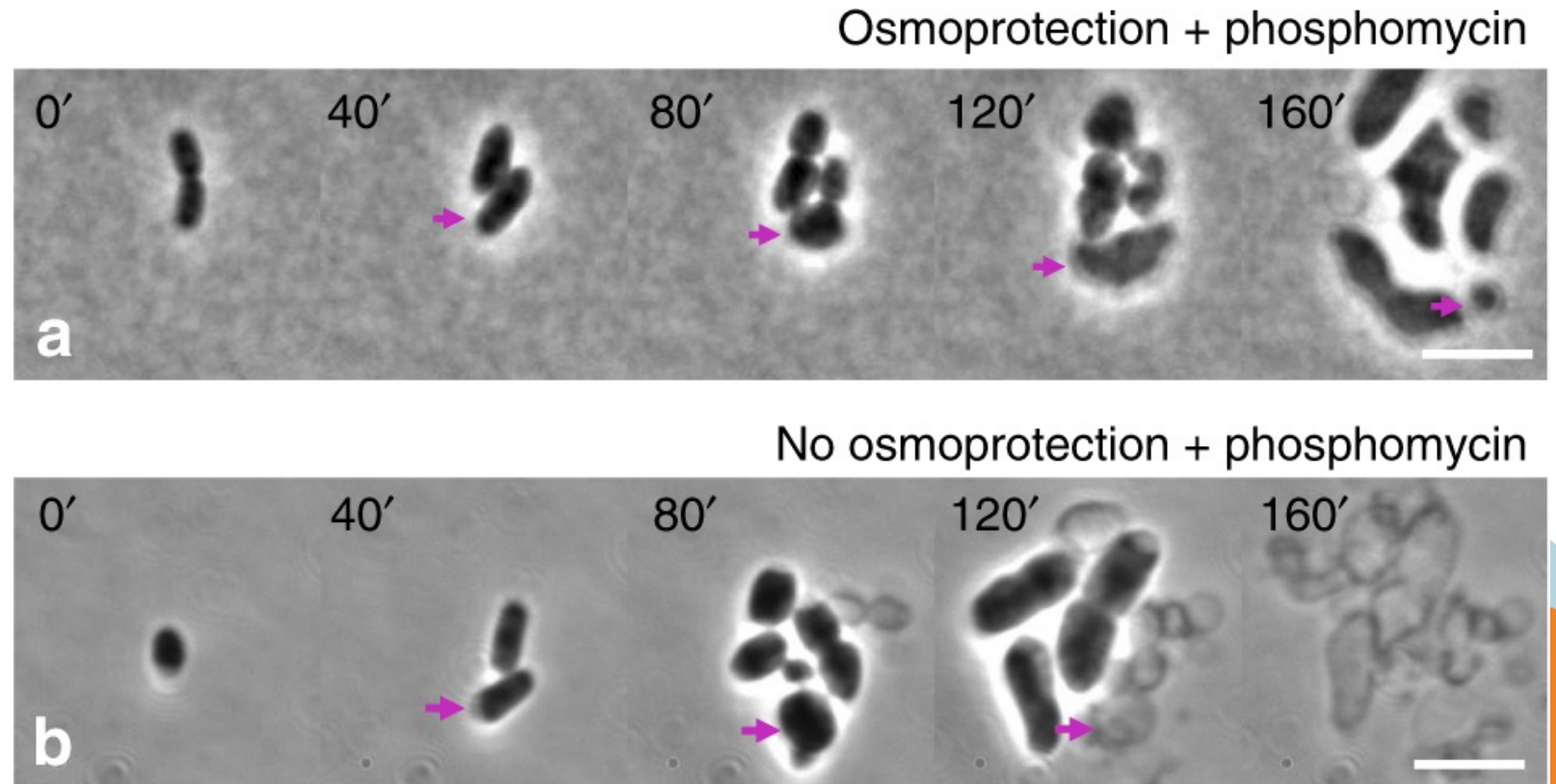
# L-Forms found in rUTI patients via fluorescence *in situ* hybridisation (FISH)

- fixed and stained **DAPI** and fluorescent DNA probe against bacterial 16S rRNA (**BAC 16S**)



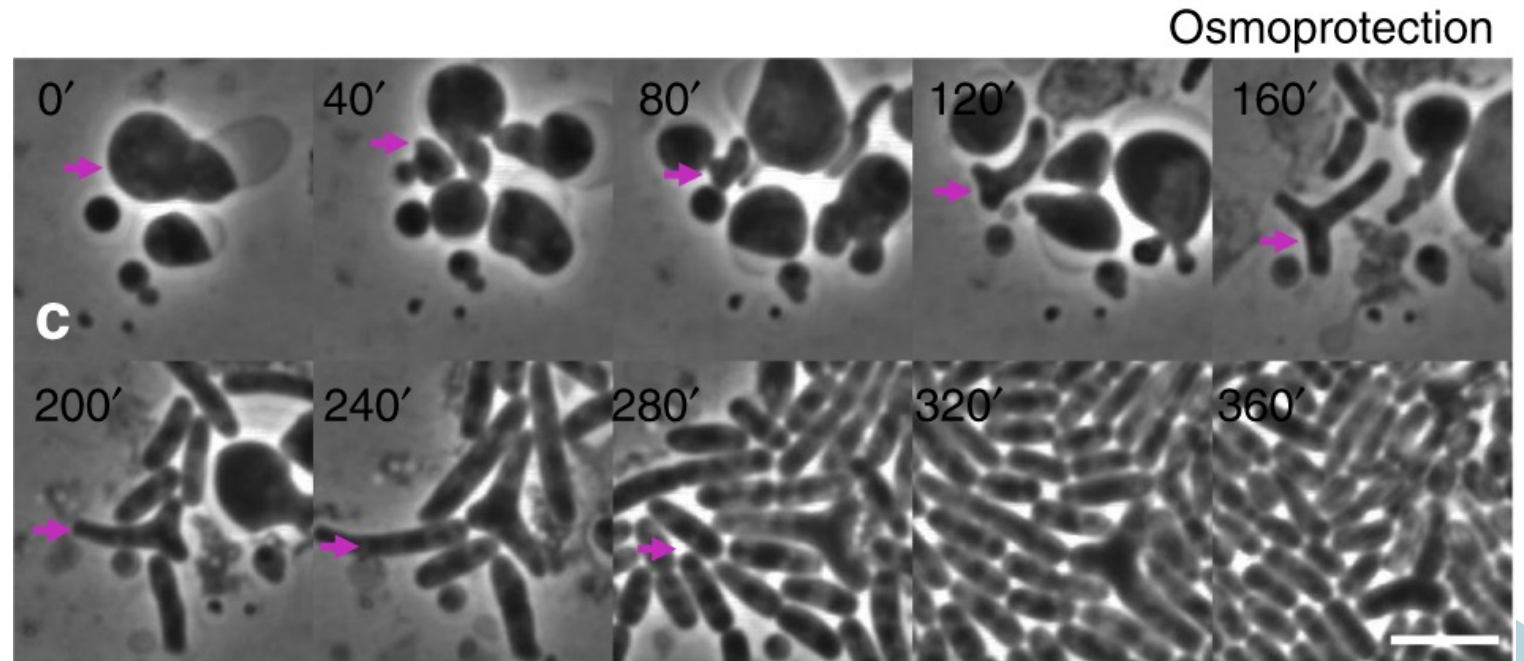
# *E. coli* switches between L-form and walled states

- *E. coli* time-lapse microscopy in presence of **phosphomycin**,
  - with and without osmoprotection

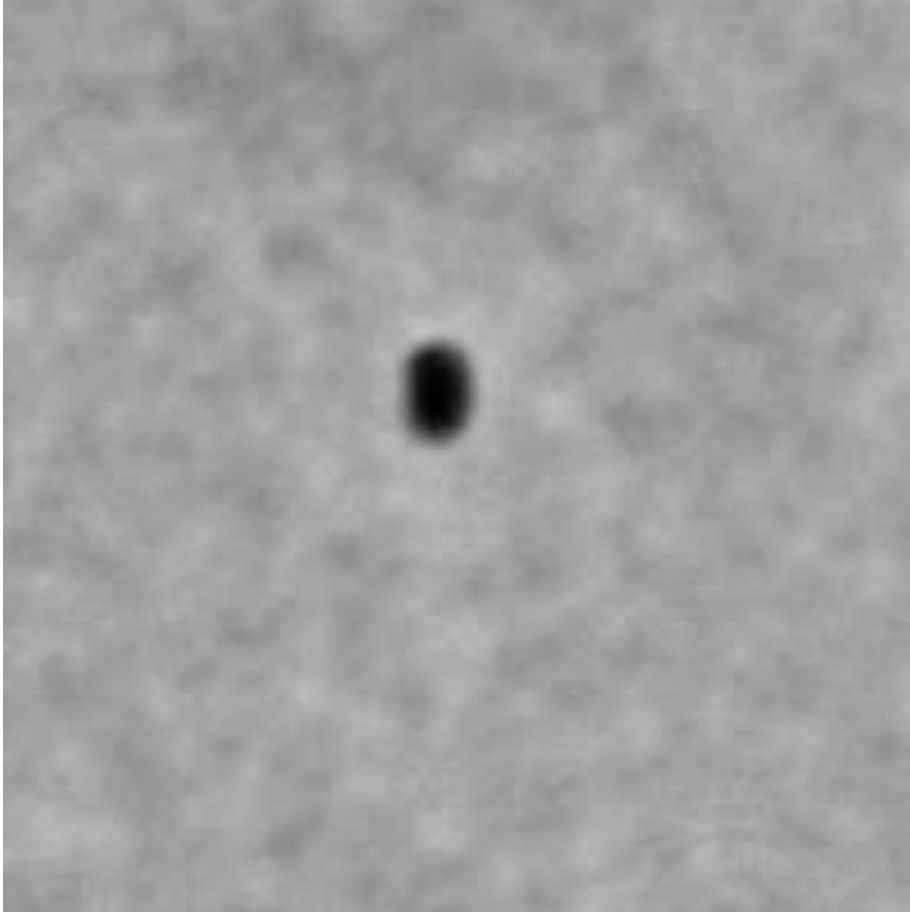


# L-form *E. coli* reverts to walled states following phosphomycin removal

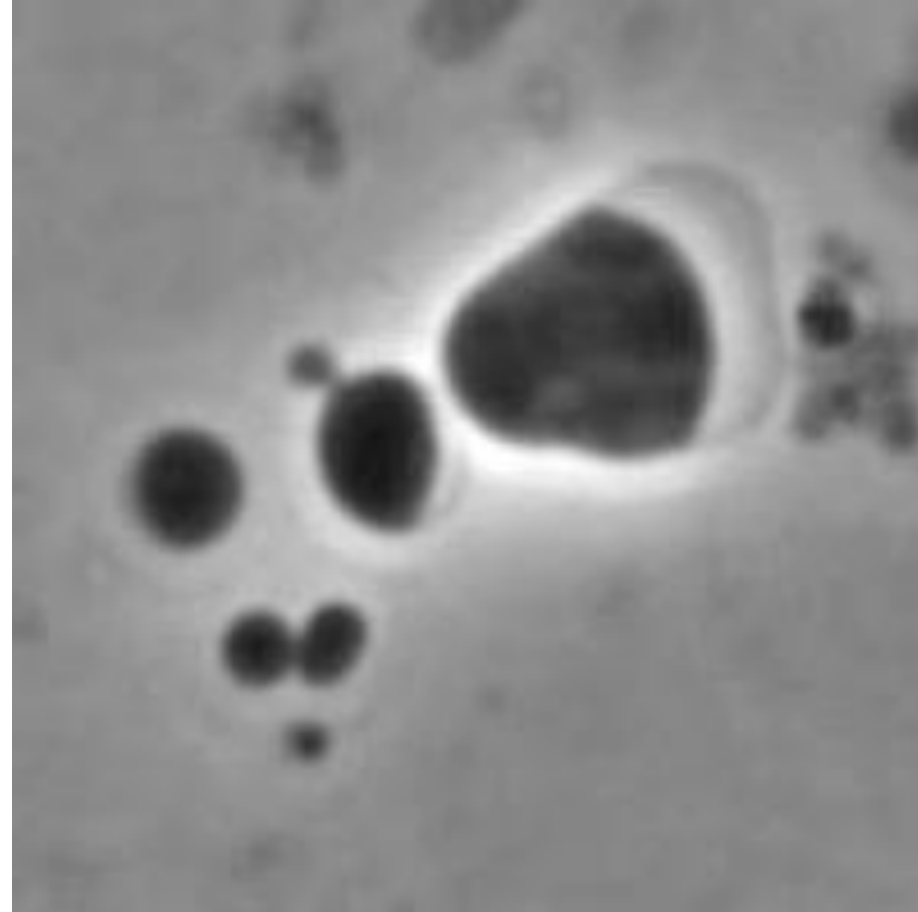
- 5 h time lapse
  - 40 min increments
- L-form *E. coli* time-lapse microscopy in removal of phosphomycin



# L-form switching time-lapse



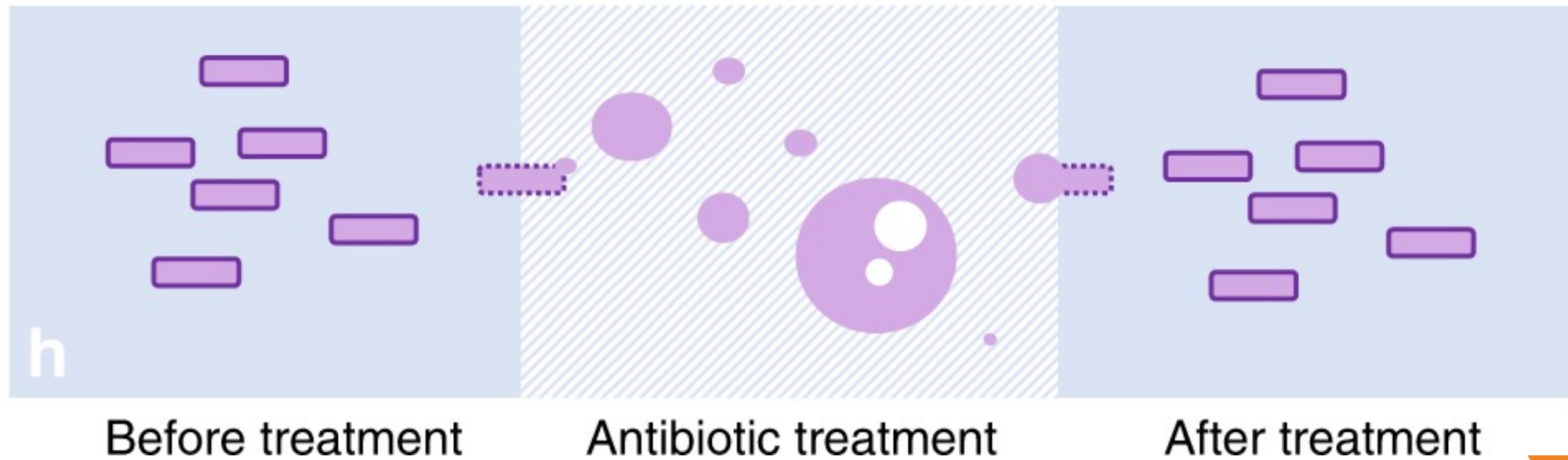
*E. coli* time-lapse microscopy in presence of phosphomycin.



*E. coli* time-lapse microscopy in absence of phosphomycin

# Summary: L-form transition allows bacteria to evade antibiotic challenge in rUTIs

- L-form switching is a clinically relevant phenomenon that may contribute to the recurrence of infection in rUTI patients

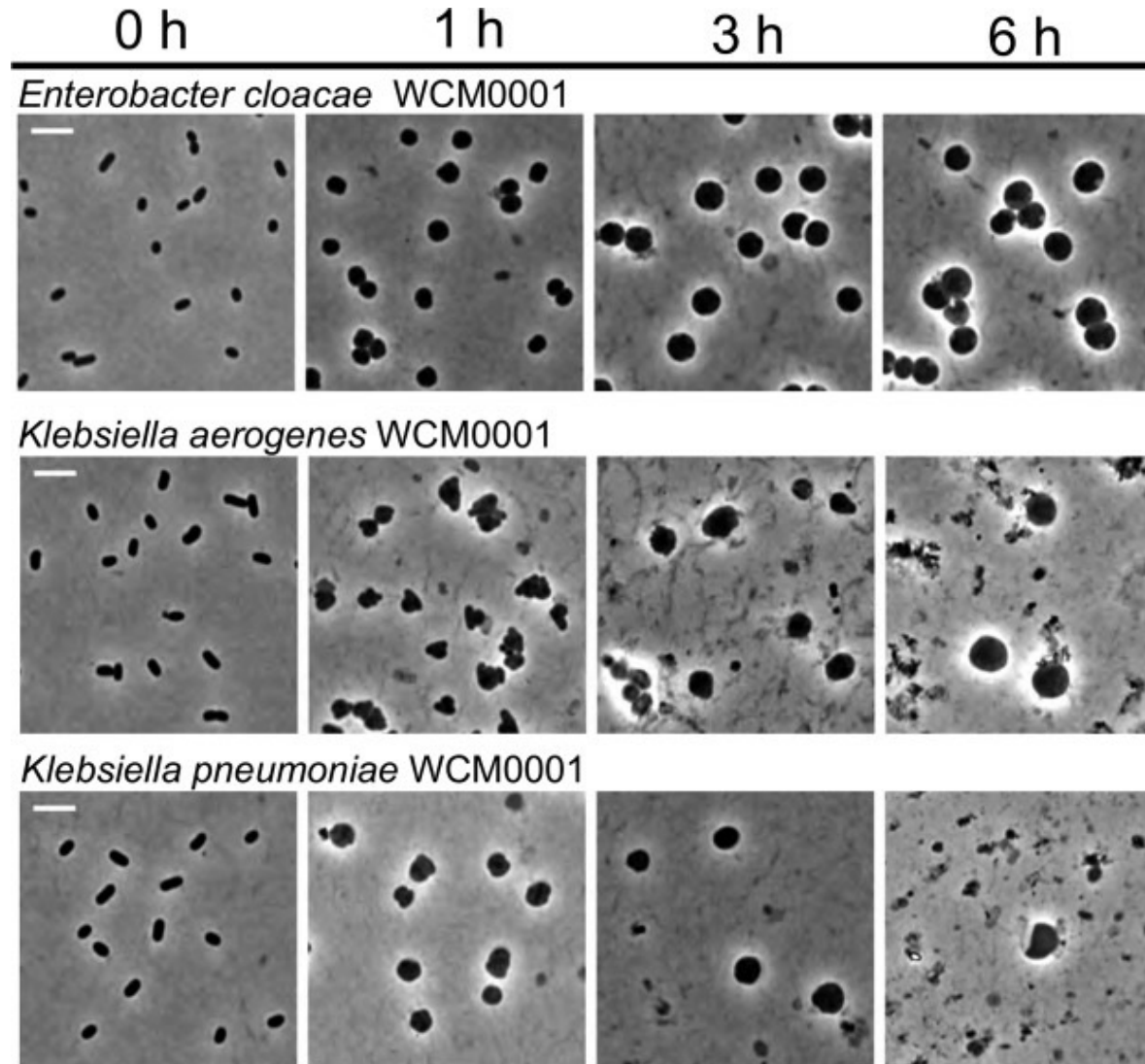


# Can other bacteria become L-forms?

Infectious agent	Example on ability to switch to L-form
<i>Klebsiella pneumoniae</i>	Cross et al., <b>Spheroplast-Mediated Carbapenem Tolerance in Gram-Negative Pathogens</b> . Antimicrob Agents Chemother. 2019 Aug 23;63(9):e00756-19. doi: 10.1128/AAC.00756-19. PMID: 31285232; PMCID: PMC6709500.
<i>Proteus mirabilis</i>	Tölg et al., <b>Dependence of induction of enterobacterial AmpC beta-lactamase on cell-wall peptidoglycan, as demonstrated in Proteus mirabilis and its wall-less protoplast L-form</b> . J Gen Microbiol. 1993 Nov;139(11):2715-22. doi: 10.1099/00221287-139-11-2715. PMID: 8277255.
<i>Pseudomonas aeruginosa</i>	White et al., <b>Cell wall characteristics of Pseudomonas aeruginosa and its carbenicillin-induced L-form</b> . Acta Biol Acad Sci Hung. 1978;29(1):67-74. PMID: 112813.
<i>Enterococcus</i> spp.	Kawai et al., <b>Crucial role for central carbon metabolism in the bacterial L-form switch and killing by <math>\beta</math>-lactam antibiotics</b> . Nat Microbiol. 2019 Oct;4(10):1716-1726. doi: 10.1038/s41564-019-0497-3. Epub 2019 Jul 8. PMID: 31285586; PMCID: PMC6755032.

# Can other bacteria become L-forms?

- Cross et al., **Spheroplast-Mediated Carbapenem Tolerance in Gram-Negative Pathogens.**
- Overnight cultures of isolates were subcultured into prewarmed BHI+ liquid medium containing 10  $\mu\text{g/ml}$  meropenem



# Antibiotics used for UTIs

Antibiotic treatment	Target
Nitrofurantoin	Bacterial ribosomal proteins
Amoxicillin-clavulanic acid	Cell wall synthesis
Phosphomycin	Cell wall synthesis
Cefuroxime	Cell wall synthesis
Levofloxacin	DNA gyrase
Ciprofloxacin	DNA gyrase
Sulfamethoxazole-trimethoprim	Folic acid pathway

First Line (Nitrofurantoin, Amoxicillin-clavulanic acid, Phosphomycin)

Second Line (Cefuroxime, Levofloxacin, Ciprofloxacin, Sulfamethoxazole-trimethoprim)

- resistance of UPEC to antimicrobial agent ranges from 14.6% to 60%
- Antibiotic sensitive bacteria could L-form switch and persist
  - Unable to be found by current routine testing
- Understanding the mechanism of L-form switching can inform clinician choice of treatment



# Conclusions and future directions

L-forms provide a source of bacterial survivors during cell wall specific antibiotics

- Continue proliferating during treatment
- Revert to walled state

L-form switching may be an underappreciated mechanism of antibiotic tolerance in chronic infections

Further understanding of L-forms may inform clinicians on better choices of treatment

Further understanding of mechanisms of L-form switching may allow development of a rapid test



Thank you for listening.  
Any questions?

# Supplementary slides

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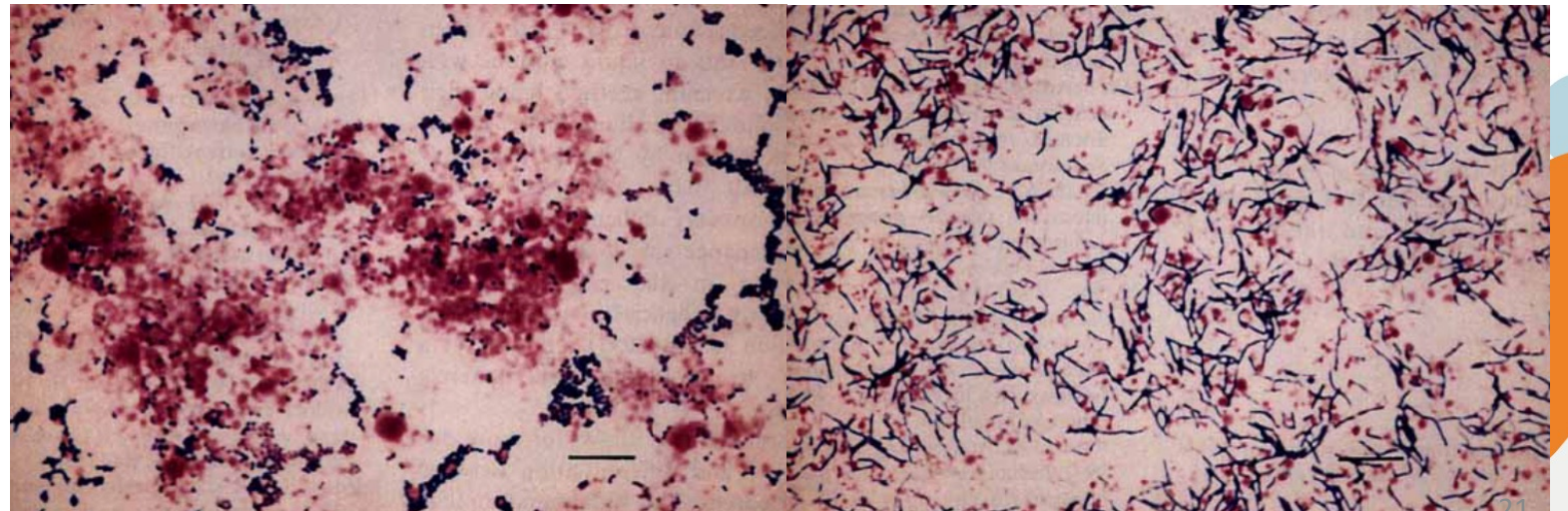
# Methods for identifying L-forms?

- Modified Gram stain
- Requires culturing with L-form media before fixing and staining
- L-forms are red due to counterstain

**A novel method for differentiating L-form bacteria from their parental form using the Hucker Gram staining technique**

E.J. ALLAN, J. JASS\*, L.E. PHILLIPS\*, J.W. COSTERTON† & H.M. LAPPIN-SCOTT\* *Department of Agriculture, University of Aberdeen, 581 King Street, Aberdeen, AB9 1UD, \*Department of Biological Sciences, Hatherly Laboratories, University of Exeter, EX4 4PS, UK and †Department of Biological Sciences, University of Calgary, Calgary, Alberta, T2N 1N4, Canada*

*DST/10: received 3 June 1992 and accepted 15 June 1992*

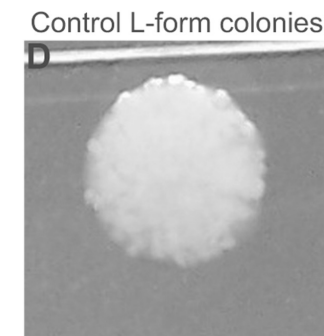
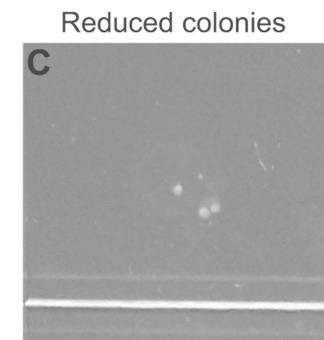
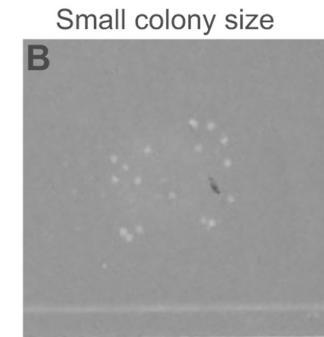
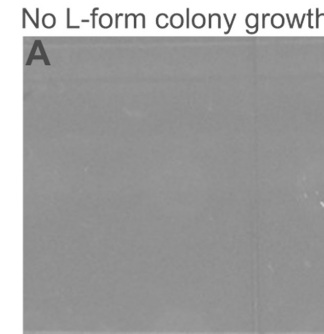


(Allan et al., 1992, *Letters in Applied Microbiolo.*)

# Methods for identifying L-forms?

- Glover et al., (2009) Insights into the **Molecular Basis of L-Form Formation and Survival in *Escherichia coli***
- **csB**, **ruvA**, **fur**, and **smpA** that are essential in the formation or survival of L-form colonies
- Pathways involved:
  - Cell envelope stress
  - DNA repair
  - Iron homeostasis
  - Drug efflux/ABC transporters
  - Outer membrane biogenesis

(Glover et al., 2009, *PLOS one*)



## Group 1 mutants (24)

*rscC*, *rscB*, *rscF*, *cpsB*, *wcaA*, *wcaF*, *wcaI*, *wza*, *wzb*, *wzc*, *wzx*, *gmd*, *galU*, *manA*, *smpA*, *yfgL*, *ruvA*, *recG*, *fur*, *yhdP*, *acrA*, *acrB*, *yrbC*, *mrcB*

## Group 2 mutants (18)

*yjbG*, *dnaT*, *recB*, *recC*, *efp*, *galE*, *wcaJ*, *priA*, *fes*, *dedD*, *zwf*, *ubiE*, *ubiF*, *lpxM*, *yraO*, *smpB*, *damX*, *yhcM*

## Group 3 mutants (10)

*wcaL*, *fcl*, *vacJ*, *yrbD*, *yrbE*, *rffD*, *ompA*, *recA*, *fis*, *cpxA*

BW25113 parent

# Methods for identifying L-forms?

- **L-form switching in *B. subtilis***
  - Two point mutations are needed for L-form growth in *B. subtilis*
1. *rodA* -> induces L-form, unable to grow
    - Peptidoglycan glycosyltransferase RodA
    - **cell wall elongation** and the **maintenance of the rod cell shape**
  2. *ispA* -> prevent lysis of L-form during growth
    - catalyzes the formation **farnesyl pyrophosphate**
    - required for synthesis of the precursors for **peptidoglycan** (lipid II) and **wall teichoic acid**

Cell

Volume 152, Issue 5, 28 February 2013, Pages 997-1007



Article

## Excess Membrane Synthesis Drives a Primitive Mode of Cell Proliferation

Romain Mercier<sup>1, 2</sup>, Yoshikazu Kawai<sup>1, 2</sup>, Jeff Errington<sup>1</sup>  

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Current Biology

Volume 25, Issue 12, 15 June 2015, Pages 1613-1618



Report

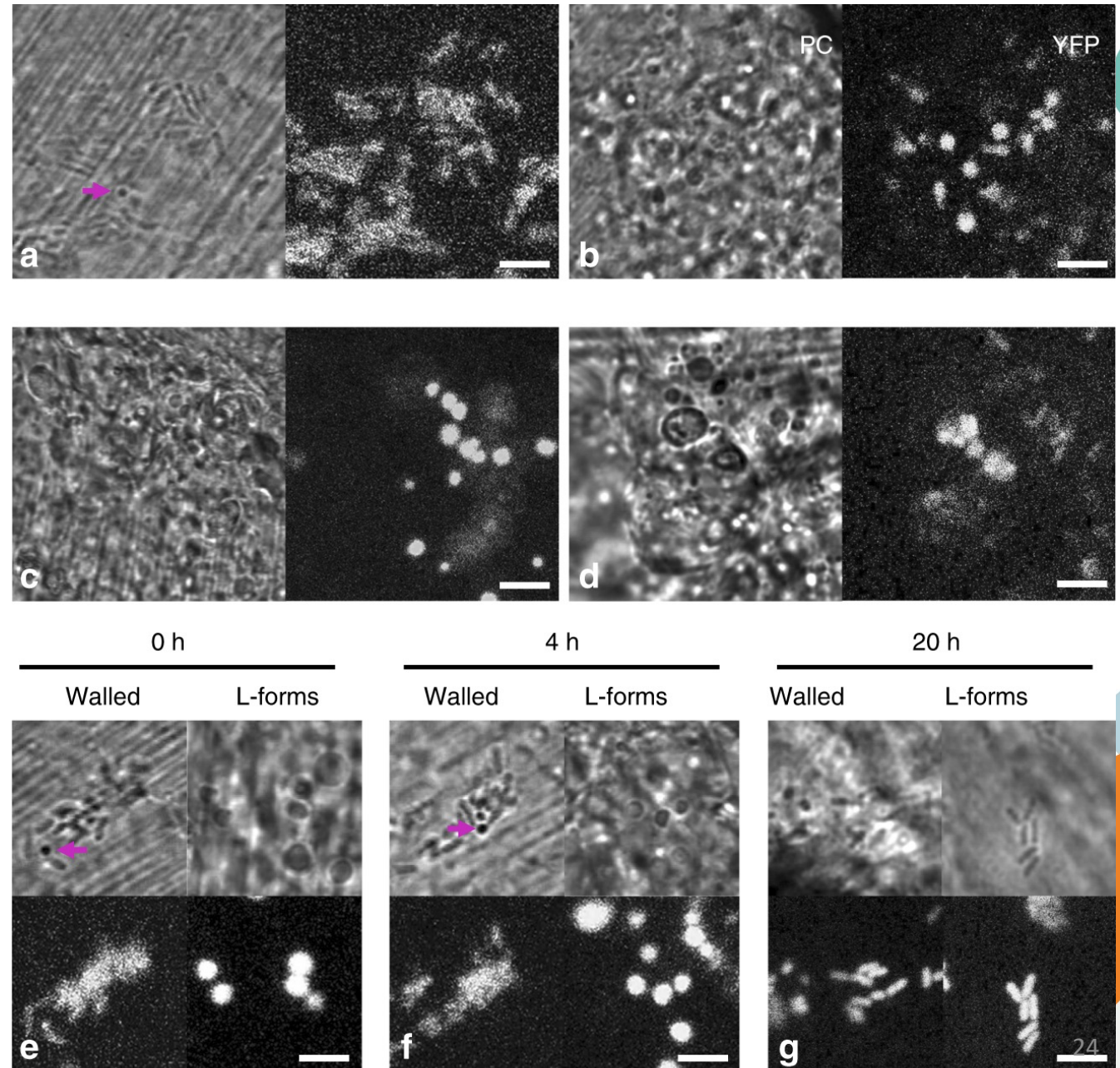
## Cell Growth of Wall-Free L-Form Bacteria Is Limited by Oxidative Damage

Yoshikazu Kawai<sup>1, 4</sup>  , Romain Mercier<sup>1, 4</sup>, Ling Juan Wu<sup>1</sup>, Patricia Domínguez-Cuevas<sup>2</sup>, Taku Oshima<sup>3</sup>, Jeff Errington<sup>1</sup>  

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# In vivo model

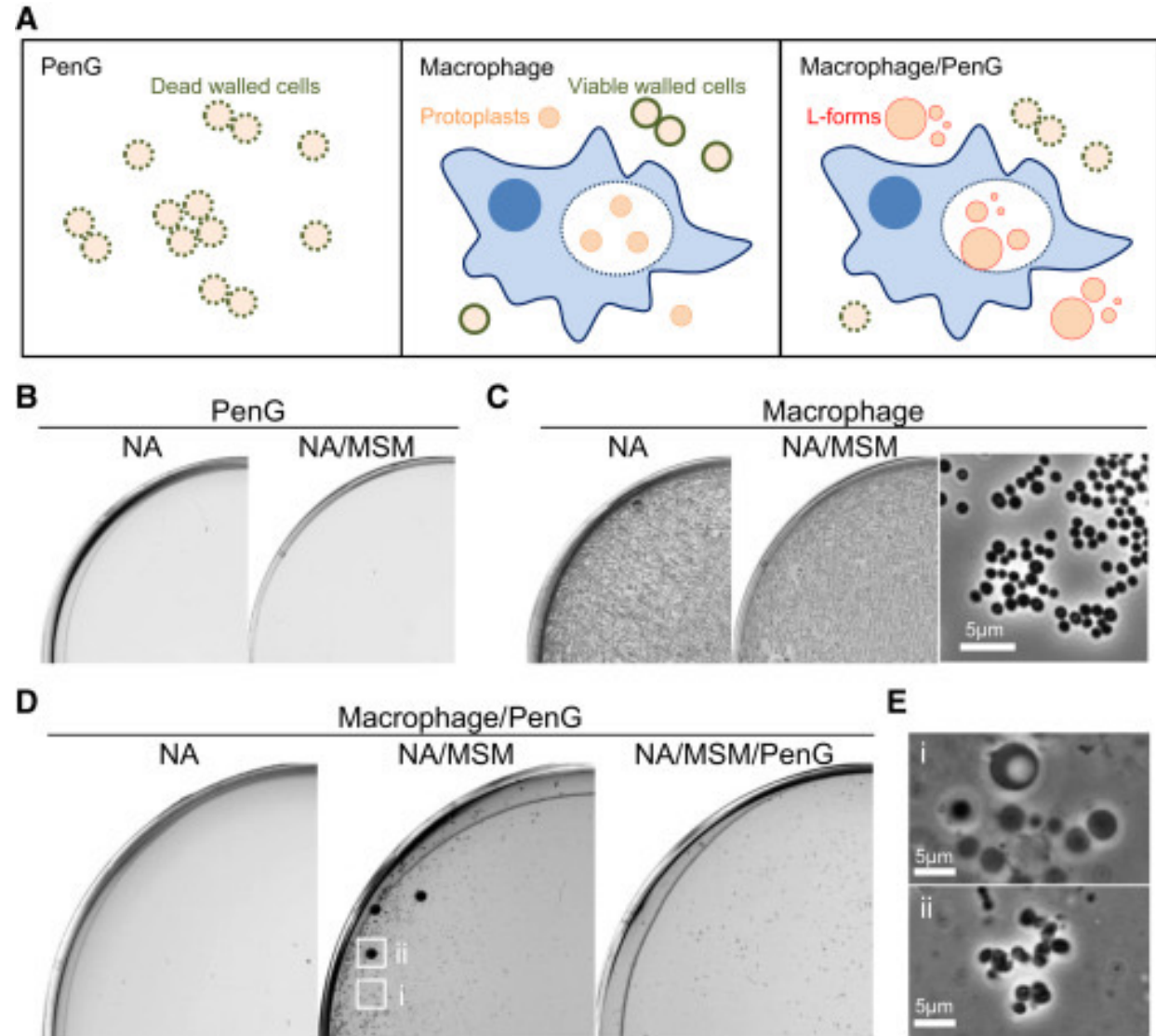
- Transparent zebrafish larvae
- A-D: L-form *E. coli* injected *in vivo* with/without antb
  - L in antb
  - L->N without antb
- E-G: Injected with walled/L-form *E. coli* without antb
  - L->N switch *in vivo*
- Homogenise embryo ☹️ and plated then counted colonies





# Other ways in which L-forms can be generated naturally

- Bacterial Cells Evade Antibiotic Action of PenG by Interacting with Macrophages
- Interaction with **lysozymes** can induce L-form switching



# Other ways in which L-forms can be generated naturally

“L-form switching is a **common response** of pathogenic *E. coli* strains to **cell wall targeting antibiotics** and that the most commonly used **lytic bacteriophages** are ineffective against them in this state”

## L-form switching confers antibiotic, phage and stress tolerance in pathogenic *Escherichia coli*

Aleksandra Petrovic Fabijan<sup>1</sup>, Muhammad Kamruzzaman<sup>1</sup>, David Martinez-Martin<sup>2,3</sup>, Carola Venturini<sup>1,4</sup>, Katarzyna Mickiewicz<sup>5</sup>, Neftali Flores-Rodriguez<sup>6</sup>, Jeff Errington<sup>5</sup>, Jonathan R. Iredell<sup>1,4,7</sup>

1. Centre for Infectious Diseases and Microbiology, Westmead Institute for Medical Research, Sydney, New South Wales, Australia
2. School of Biomedical Engineering, The University of Sydney, Sydney, New South Wales, Australia
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7. Westmead Hospital, Western Sydney Local Health District, Sydney, New South Wales, Australia

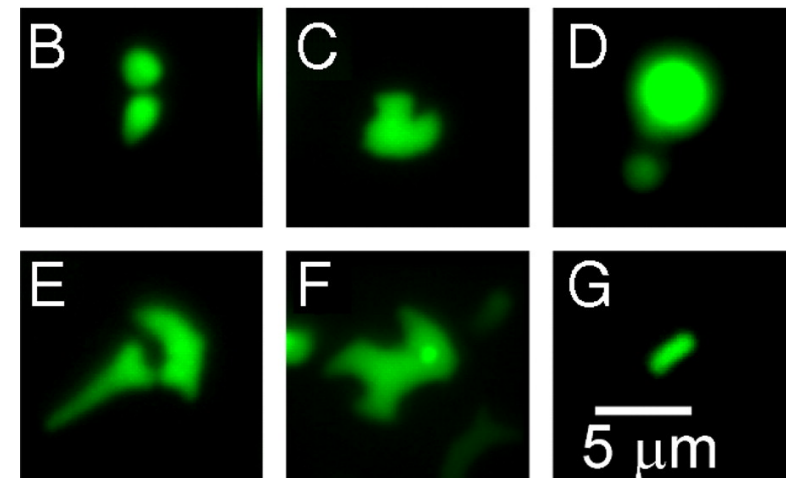
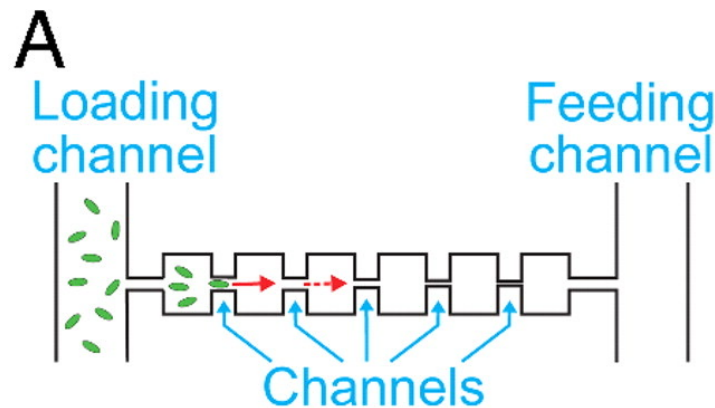
# Other ways in which L-forms can be generated naturally

- Microfluidic channels (constrictions)
- *E. coli* L-forms are derived after penetrating channels with a width that is smaller than their diameter
- L-form switching appears to be a stress response

## Bacterial growth and motility in sub-micron constrictions

Jaan Männik, Rosalie Driessen, Peter Galajda, Juan E. Keymer, and Cees Dekker  
+ [See all authors and affiliations](#)

PNAS September 1, 2009 106 (35) 14861-14866; <https://doi.org/10.1073/pnas.0907542106>



(Männik et al., 2009, PNAS)

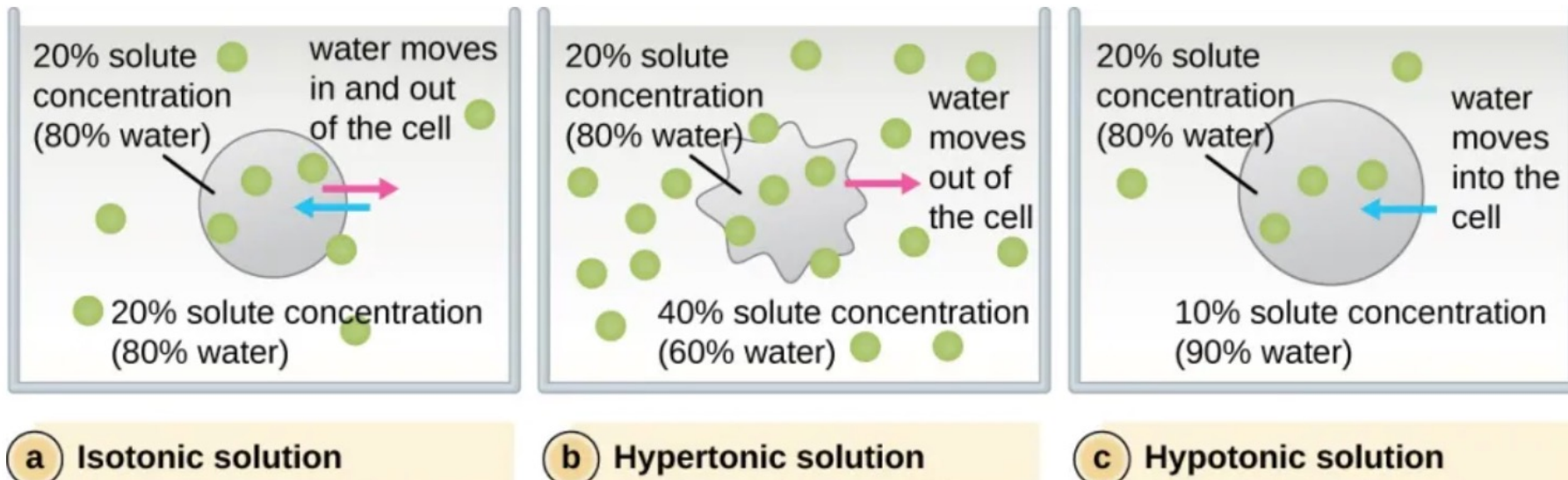
# Other strains of pathogenic E. coli isolated from other specimen types display L-form switching due to stress from meropenem and lytic phages

E. coli strain ID	Specimen	ST (clade)	Antimicrobial resistance genes	Meropenem MIC (µg/ml)		Specific phages			
				WT	Revertant	Eco2	Eco6	Eco11	Eco12
SYD045	Urine	ST1193	<i>bla<sub>CTX-M-14a</sub></i> , <i>bla<sub>TEM-1b</sub></i> , <i>aac(3)-Ild</i>	<0.25	ND	Dark Green	Grey	Grey	Dark Green
SYD252	Urine	ST131 (B)	<i>bla<sub>CTX-M-15</sub></i> , <i>bla<sub>TEM-1b</sub></i> , <i>aac(3)-Ild</i> , <i>dfrA17-aadA5</i> , <i>sul1</i> , <i>mph(A)</i> , <i>sul2</i> , <i>strAB</i> , <i>tet(A)</i>	0.031	0.031	Grey	Grey	Dark Green	Grey
SYD449	Blood culture	ST131 (A)	None	0.031	0.031	Grey	Grey	Grey	Yellow
SYD214	Urine	ST648	<i>bla<sub>TEM-1b</sub></i> , <i>aac(3)-Ile</i>	<0.25	ND	Grey	Grey	Grey	Yellow
SYD402	Urine	ST73	<i>bla<sub>TEM-1b</sub></i> , <i>sul2</i>	0.031	0.015	Light Green	Grey	Grey	Light Green
SYD009	Blood culture	ST95	<i>bla<sub>DHA-1</sub></i> , <i>qnrB4</i> , <i>dfrA17</i> , <i>sul1</i> , <i>mph(A)</i> , <i>sul2</i> , <i>strAB</i> , <i>tet(B)</i>	0.015	0.015	Yellow	Grey	Grey	Yellow
SYD074	Blood culture	ST58	<i>bla<sub>TEM-1b</sub></i> , <i>dfrA5</i> , <i>strAB</i> , <i>sul2</i> , <i>tet(A)</i>	0.015	0.015	Grey	Grey	Grey	Grey
SYD066	Urine	ST405	<i>bla<sub>CTX-M-15</sub></i> , <i>aac(3)-Ile</i> , <i>dfrA17-aadA5</i> , <i>sul1</i> , <i>mph(A)</i> , <i>tet(B)</i>	<0.25	ND	Dark Green	Grey	Grey	Dark Green
SYD259	Urine	ST998	<i>bla<sub>CTX-M-15</sub></i> , <i>bla<sub>TEM-1b</sub></i> , <i>bla<sub>OXA-1</sub></i> , <i>aac(6)-Ib-cr</i> , <i>aac(3)-Ile</i> , <i>aphA1</i> , <i>dfrA5</i> , <i>dfrA17-aadA5</i> , <i>sul1</i> , <i>mph(A)</i> , <i>sul2</i> , <i>floR</i>	0.015	0.015	Grey	Grey	Grey	Grey
SYD001	Urine	ST38	<i>bla<sub>CTX-M-27</sub></i> , <i>dfrA17-aadA5</i> , <i>sul1</i> , <i>mph(A)</i> , <i>sul2</i> , <i>strAB</i> , <i>tet(A)</i>	0.031	0.031	Grey	Grey	Grey	Grey
SYD421	Urine	ST349	<i>bla<sub>TEM-1b</sub></i> , <i>dfrA14</i> , <i>sul2</i> , <i>strAB</i>	0.031	0.031	Yellow	Grey	Grey	Grey
JIE4039	Urine	ST963	<i>bla<sub>CMY-2</sub></i>	0.031	0.063	Grey	Light Green	Grey	Grey
B36	Blood culture	ST131 (C)	<i>bla<sub>CTX-M-15</sub></i> , <i>bla<sub>TEM-1b</sub></i> , <i>bla<sub>OXA-1</sub></i> , <i>dfrA17-aadA5</i> , <i>sul1</i> , <i>mph(A)</i> , <i>sul2</i> , <i>strAB</i> , <i>tet(A)</i>	0.031	0.031	Grey	Grey	Grey	Light Green
J53	Stool	ST10	None	<0.25	ND	Grey	Dark Green	Grey	Yellow
WH62	Stool	ST127	None	0.063	0.063	Dark Green	Grey	Grey	Dark Green

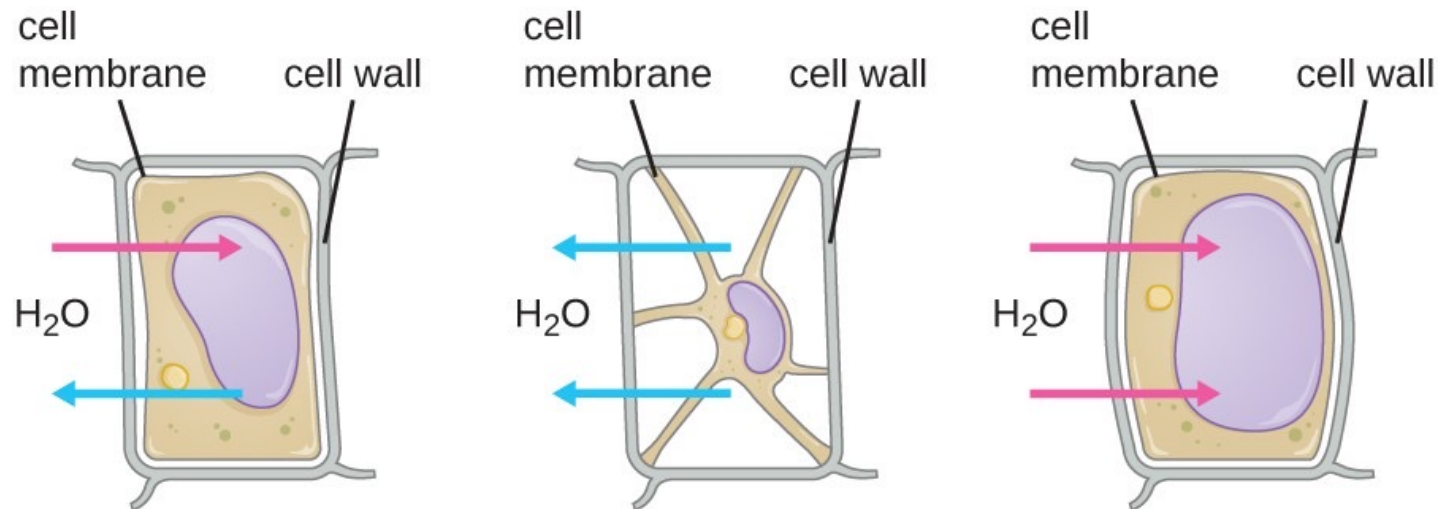
\*MIC=minimal inhibitory concentration; ND=not determine; Coloured cells represent phage activity on CWB lawn: dark green=clear plaques, light green=slightly turbid plaques, yellow=turbid plaques and grey=no activity.

# Routine microbiological media is hypotonic and does not support L-form growth

L-form



N-form



# L-forms provide a route for antibiotic evasion

- patient UTI343
- treated with **phosphomycin** (donation 6)
- Significant viable bacterial  $>1 \times 10^5$
- **UPEC strain ST144**

